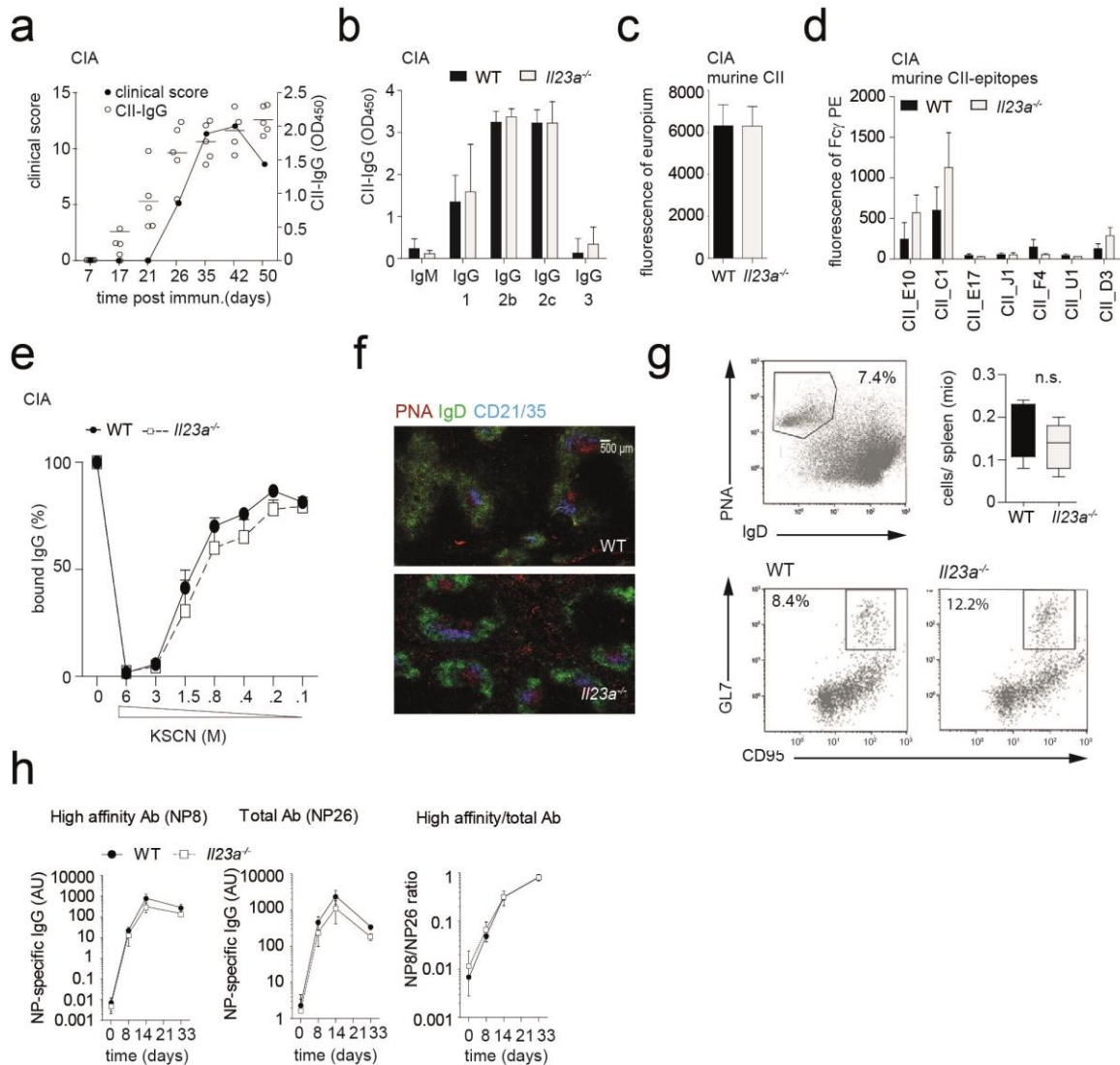


Supplementary Figure 1

Il23a^{-/-} mice develop regular arthritis induced by the transfer of K/BxN serum.

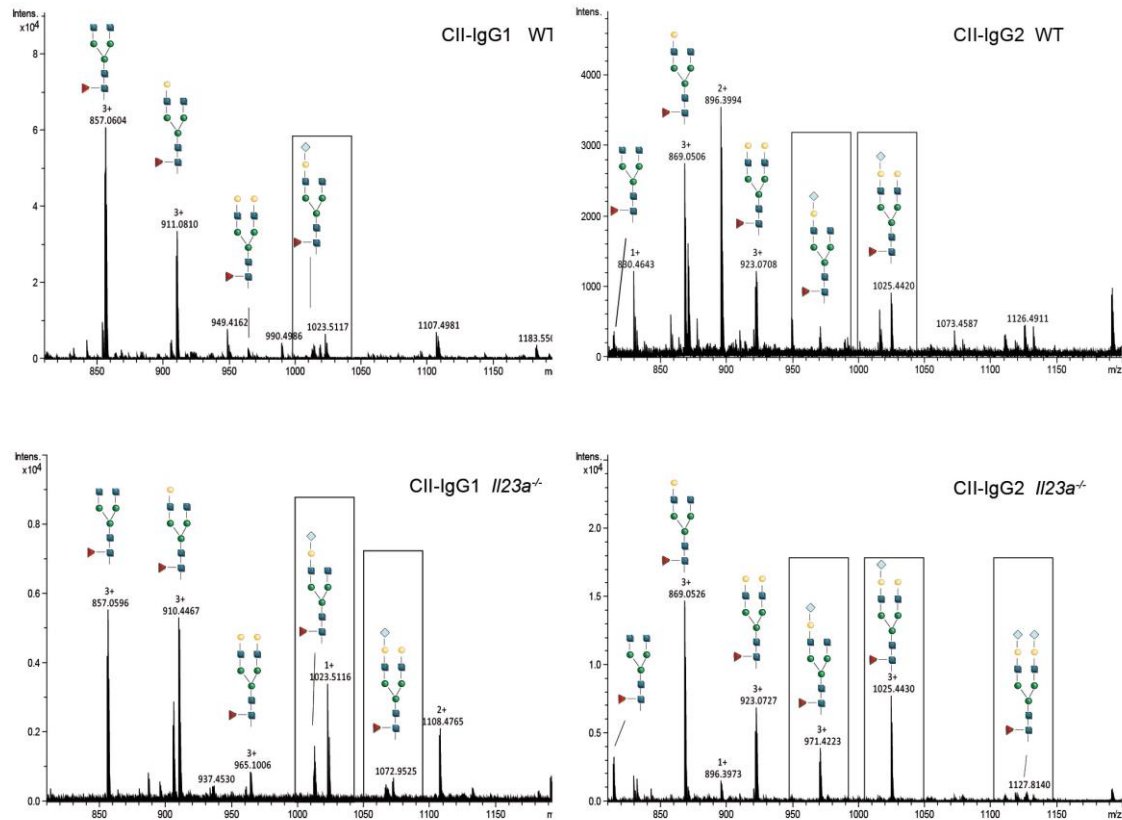
(a) Clinical scoring of arthritis in wild type (WT) and *Il23a*^{-/-} mice that received serum from arthritic K/BxN mice. Error bars represent SEM *P<0.05; **P<0.01; ***P<0.001; Student's t test.



Supplementary Figure 2

Humoral immune response in wild-type and *Il23a*^{-/-} mice during CIA.

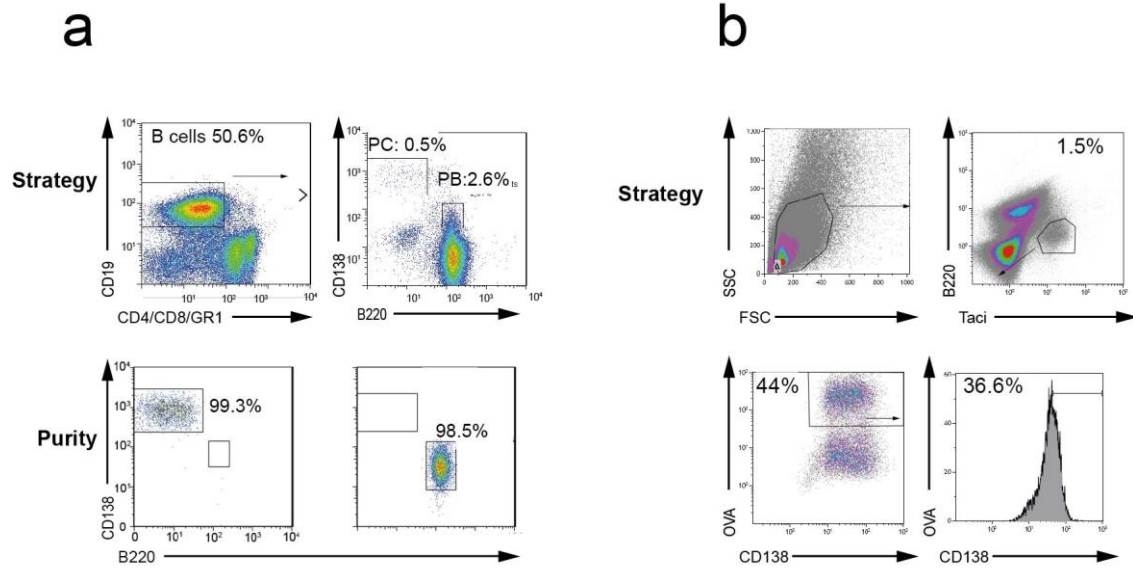
(a-d) Analysis of levels of (a) collagen type II (CII)-specific IgG, (b) CII-specific IgG subsets, (c) IgG specifically directed against murine CII (d) and IgG detecting different murine CII epitopes in the sera of WT and *Il23a*^{-/-} mice at day 50 after induction of collagen-induced arthritis (CIA). (e) KSCN titration assay for the determination of the binding affinity of CII-specific IgG in WT and *Il23a*^{-/-} mice. (f,g) Analysis of the appearance of (f) germinal centers and the appearance of germinal center (CD19 pre-gated) B cells (g) at day 26 after induction of CIA in WT and *Il23a*^{-/-} mice. (h) Measurement of the levels of regular and high affinity antibodies in response to a T cell dependent immunization with 4-hydroxy-3-nitrophenylacetyl (NP) coupled to chicken g-globulin (NP-CGG) in WT and *Il23a*^{-/-} mice. Error bars represent SEM**P*<0.05; ***P*<0.01; ****P*<0.001; Student's *t* test.



Supplementary Figure 3

Analysis of the glycostructure of CII-specific IgG by mass spectrometry.

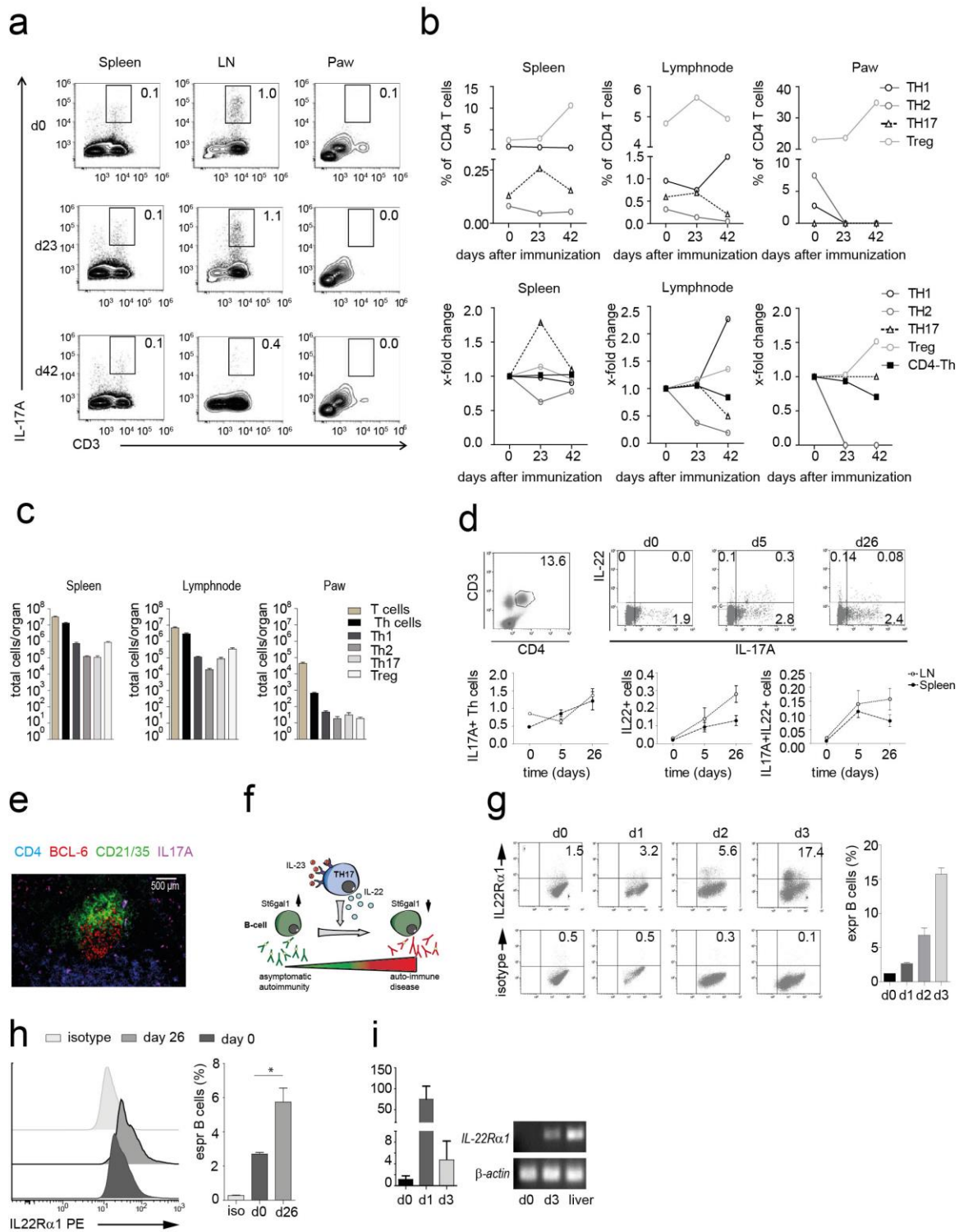
Representative mass spectra illustrating IL-23-dependent changes of the glycosylation at Asn-297 of the indicated IgG subclasses of collagen type II-specific IgG isolated from the sera of WT and *//23a^{-/-}* mice at day 50 after induction of CIA.



Supplementary Figure 4

Identification of plasma cells and plasmablasts.

(a) Sorting strategy and sorting purity analysis of splenic plasma cells ($CD19^+CD138^+B220^-$) and plasmablasts ($CD19^+CD138^{low}B220^+$) for mRNA-analysis. (b) Gating strategy for the flow cytometric analysis of St6gal1 protein expression in splenic Ova-specific plasmacells and plasmacells isolated during collagen-induced arthritis. Plasmacells were gated to be $B220^{low}CD138^+Taci^+$ and Ova^+ , respectively.

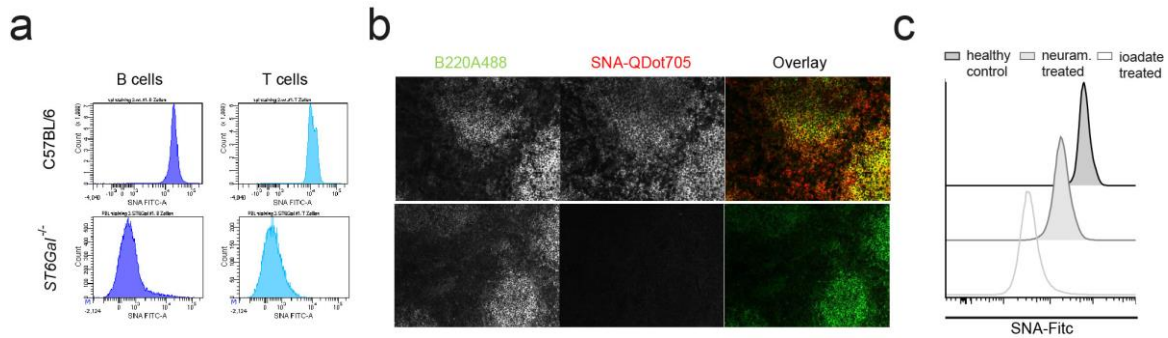


Supplementary Figure 5

Characterization of the T_H17 response during CIA.

(a) Flow-cytometric analysis of the content of IL-17-expressing $CD3^+$ T cells in spleen, inguinal lymph nodes (LN) and paws of wild-type (WT) DBA mice at indicated time points during the course of collagen-induced arthritis (CIA). (b) Mean percentage and fold change of the numbers as well as (c) total cells/organ of indicated T cell subsets in spleen, LN and

paws of WT DBA/1 mice at indicated time points after induction of CIA. CD4⁺CD3⁺ T cells were subcategorized into IFN γ -expressing Th1 cells, IL-4-expressing Th2 cells, IL-17-expressing T_H17 cells and FoxP3-expressing Treg cells. **(d)** Frequency of IL-17-positive, IL-22-positive and IL-17/IL-22 double-positive T cells in spleens and lymph nodes of WT mice at the indicated time points after induction of CIA. **(e)** Identification of CD4⁺Bcl-6⁺IL17⁺ T cells within germinal centers of the spleen 26 days after induction of CIA **(f)** Proposed model of the IL23/Th17-mediated control of autoantibody activity. **(g)** Flow cytometry-based quantification of the expression of the IL-22 receptor (IL22R α 1) on CD19⁺B220⁺ B cells that were differentiated into plasmablasts by incubation with LPS (5 μ g/ml). **(h)** Quantification of IL-22 receptor (IL-22R α 1) surface expression on splenic CD19⁺B220⁺ B cells at day 0 and day 26 after induction of CIA. **(i)** mRNA expression of IL22Ra1 in in vitro differentiating plasmacells. Error bars represent SEM*P < 0.05, **P < 0.01, ***P < 0.001; Student's t test. Error bars represent SEM*P < 0.05, **P < 0.01, ***P < 0.001; Student's t test.



Supplementary Figure 6

Evaluation of the specificity of SNA lectin surface staining for determination of cellular St6gal1 activity.

(a) B and T cells of wild-type (WT) and *St6gal1*^{-/-} mice were stained with a FITC-coupled SNA-lectin to determine the levels of surface sialic acid. (b) Immunofluorescence microscopy of spleens of WT and *St6gal1*^{-/-} mice determining the levels of sialic acid on B220⁺ B cells by co-staining with an antibody against B220 (A488, green) and a SNA lectin (QDot705, red). (c) Human CD19⁺ B cells were stained with FITC-stained SNA-lectin to determine the levels of surface sialic acid. B cells were pre-treated with neuraminidase (100 mU, 1h at 37°C) or iodate (2mM, 1h at 4°C) to remove sialic acid where indicated. Error bars represent SEM *P < 0.05, **P < 0.01, ***P < 0.001; Student's t test.